

# G.I. Procedures

## Scope Culture Protocol

### Handbook



*\*Testing requires two individuals.*

## Step 1 Broth

1. Contact Memorial Hospital Epidemiology Laboratory two or three days before your scope testing. This is located on the 5<sup>th</sup> floor or call **919-843-0867**.
2. Order three broth containers per scope.
3. Bring cooler carrying container to keep broth containers cold.
4. Each container returned after testing should have 20 cc or more in order to be properly tested.
5. When testing is completed immediately deliver all specimens back to the Memorial Hospital Epidemiology Laboratory.



## Step 2 Supplies Needed

1. Two sterile SRI gowns.
2. Two pairs sterile gloves and bonnets.
3. One sterile cover or sterile gown.
4. Two 60 ml Catheter tip syringes.
5. Two 20 ml syringes.
6. Two alcohol 4x4 prep pads packages.
7. Two 3" piece sterile clear tubing.
8. Two sterile suction tubing packs.
9. Two sets of 3 small sterile containers and blank labels.
10. Two sets of 2 specimen traps and blank labels.
11. Two Olympus auxiliary tubing.
12. Two masks with eye shield.
13. Two sets of each – red suction button, blue air and water button, black biopsy cap, if testing 180's scopes water resistant caps (black cap).
14. New suction canister and tubing.
15. Choose two different types of scopes (GIF, PCF, CF etc...) to test.



### Step 3 Scope Items

All buttons and black cap need to be attached to scopes before you start culture testing. Wipe all buttons and button scope ports with an alcohol wipe.

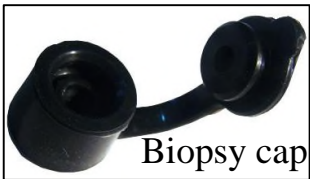
1. Suction = Red



2. Air & Water = Blue



3. The Biopsy cap needs to be on the scope before you start the culture testing.



Attach the Biopsy Cap here



4. Water Resistant Caps (Black Cap) needs to be on the 180's and lower series scopes before you start the culture testing.



Attach the Black Cap here



***\*Note the 190's scopes don't have a black cap.***

## Step 4 Scope Pre-procedure

1. Get two different scopes (GIF, PCF, CF etc...) that you want to test from the scope room (make sure not to choose from last testing).
2. Be sure you get the 3 scope buttons and water resistant cap (black cap) as well for each scope.
3. Make sure the **red** suction button goes on top (red circle around hole) and the **blue** air and water button (blue circle around hole) goes beneath it and place biopsy cap on scope.
4. Label your three specimen cups “Air”, “Water” and “Auxiliary”.
5. Label your Specimen Trap “Suction” only.
6. Label your specimen cups by recording all scope information on all specimen cups and Flexible Scope Testing log sheet.

- a. Date each specimen cup
- b. Initial each specimen cup
- c. Location: Memorial etc...
- d. Model example: GIF, CF etc...
- e. Scope Serial number
- f. Who cleaned the scope last

Flexible Scope Testing						
Date	Model#	Service	Serial#	Cleaned by	Tested by	Results
4-4-15	PEF 0180 A1	CF	3104009	SG/CL	7-3-15 CK/CL/TC/ym	OK
4-11-15	PEF 0180 A1	CF	3104009	CL/MS	7-9-15 CK/CL/TC/ym	OK
4-11-15	PEF 0180 A1	CF	3104009	CL/MS	7-9-15 CK/CL/TC/ym	OK
4-11-15	CF 180 210 367	CF	2403069	SG/PA	5-15-15 CK/CL/TC/ym	OK
4-11-15	GIF H 180 220 614	CF	220 614	SG/PA	5-15-15 CK/CL/TC/ym	OK
7/31/15	PEF 0180 A1	CF	280 7165	ms	CK/CL/TC/ym	OK
7/31	220 6 255	CF	2206255	CP	CK/CL/TC/ym	OK

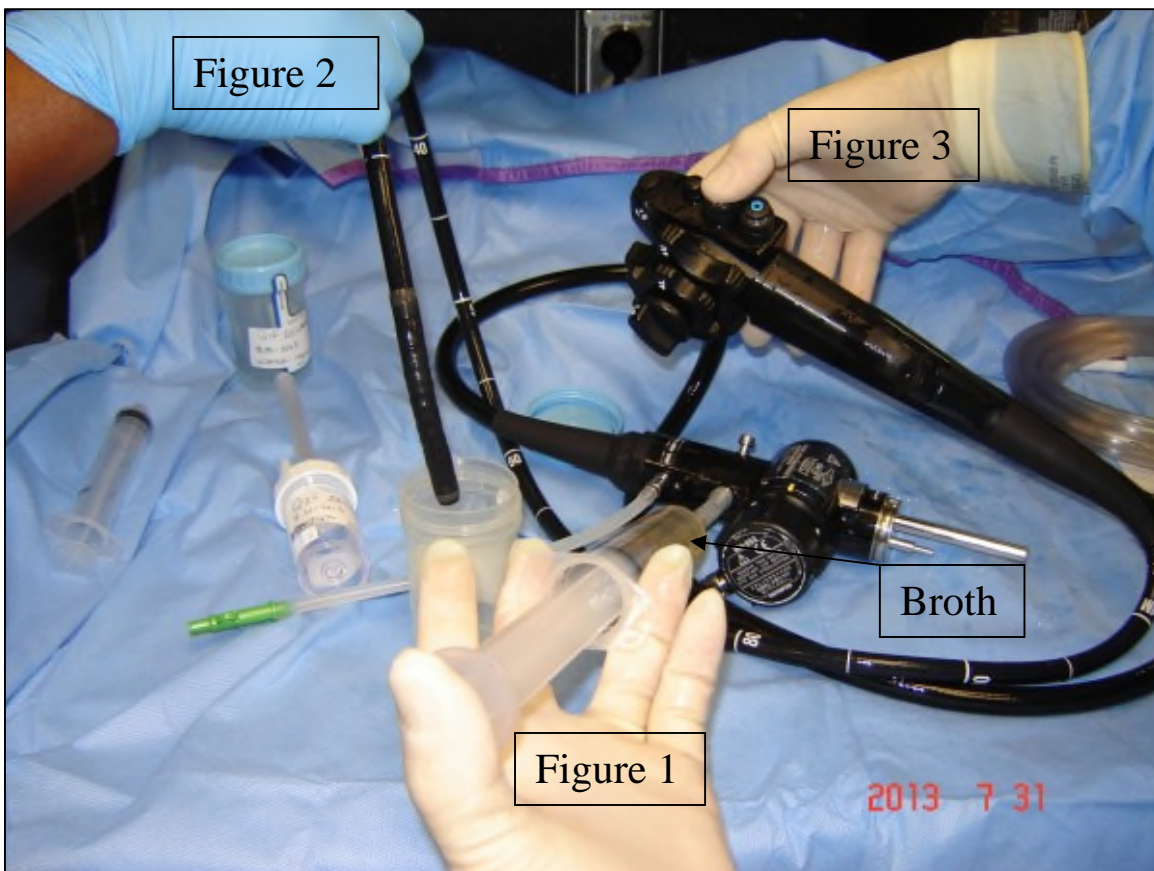
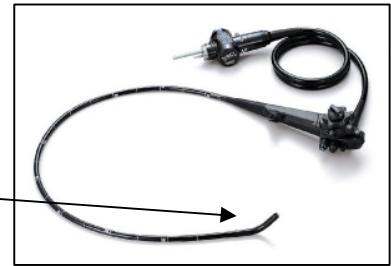
7. Suction canister and cover it with the sterile half sheet.
8. Place your first scope onto your sterile field drape.
9. Put on your Personal Protective Equipment (PPE).
10. Prepare this site by opening and placing all the items listed in **Step 2**, numbers **4 to 10** listed above onto your second sterile work area.
11. Place your tubing on your syringe.

## Step 5 Scope Culture Testing

### First – Culture the “Air” Channel

1. Place broth in the 60 ml syringe and attach syringe tubing to the silver suction nozzle (Figure 1).
2. Open appropriate specimen container.
3. Wipe all buttons and button scope ports with an alcohol wipe.
4. Wipe distal end of scope with an alcohol wipe and hold end over collection cup (Figure 2).
5. Push broth through silver suction nozzle while holding down on the **red** suction button (Figure 3).
6. Immediately close container and set aside.

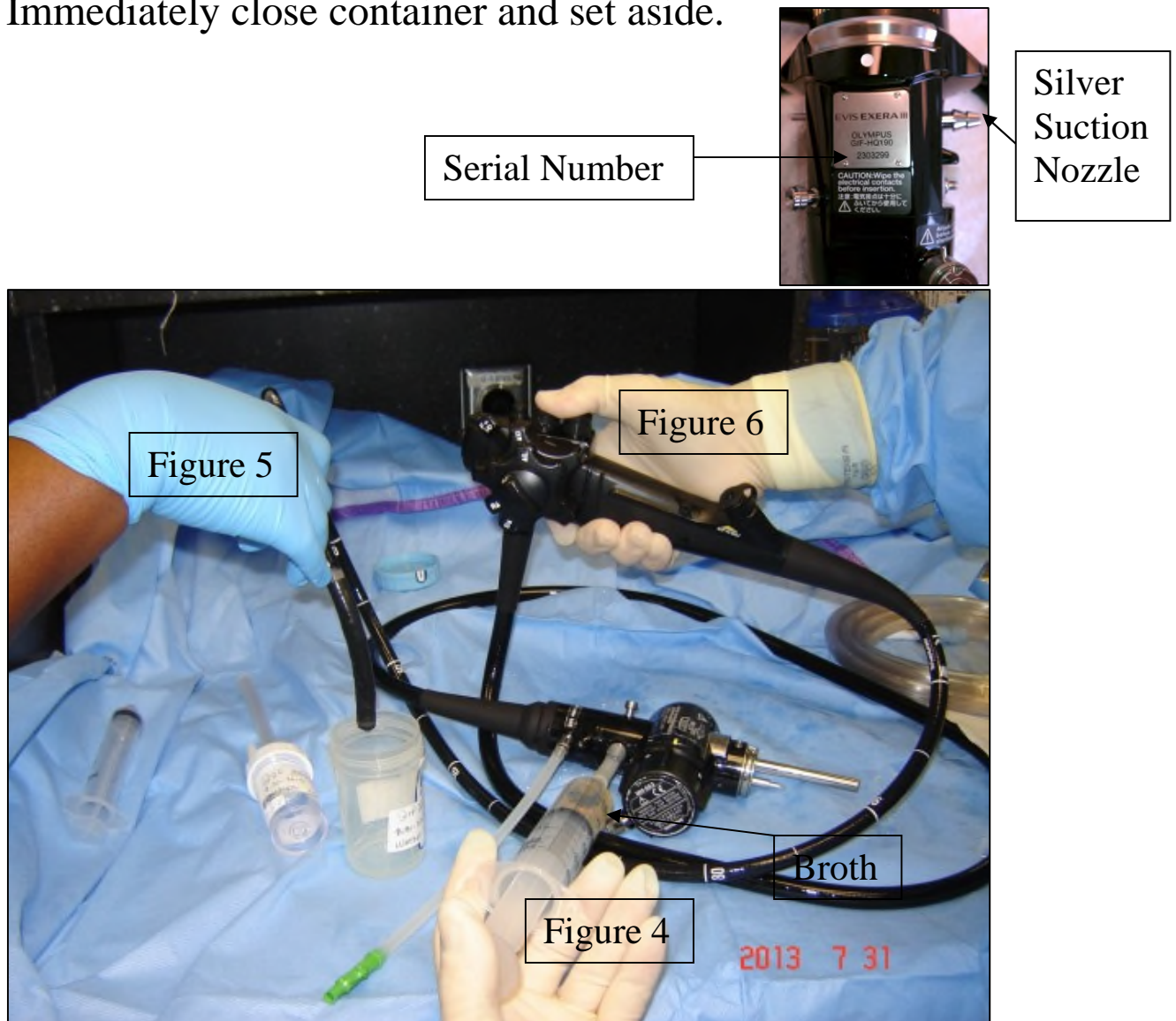
Scope Distal Section



## Step 6 Scope Culture Testing

### Second – Culture the “Water” Channel

1. Place broth in the 60 ml syringe and attach syringe tubing to the silver suction nozzle (Figure 4).
2. Open appropriate specimen container.
3. Wipe distal end of scope with an alcohol wipe and hold end over collection cup (Figure 5).
4. Push broth through silver suction nozzle while holding down on both the **red** suction button and the **blue** air and water buttons (Figure 6).
5. Immediately close container and set aside.

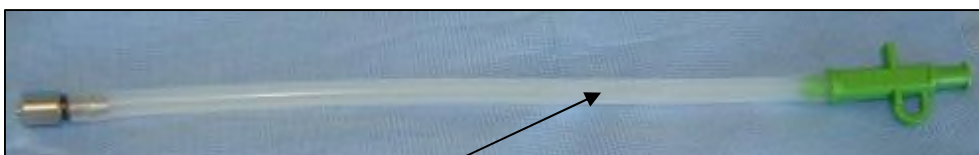
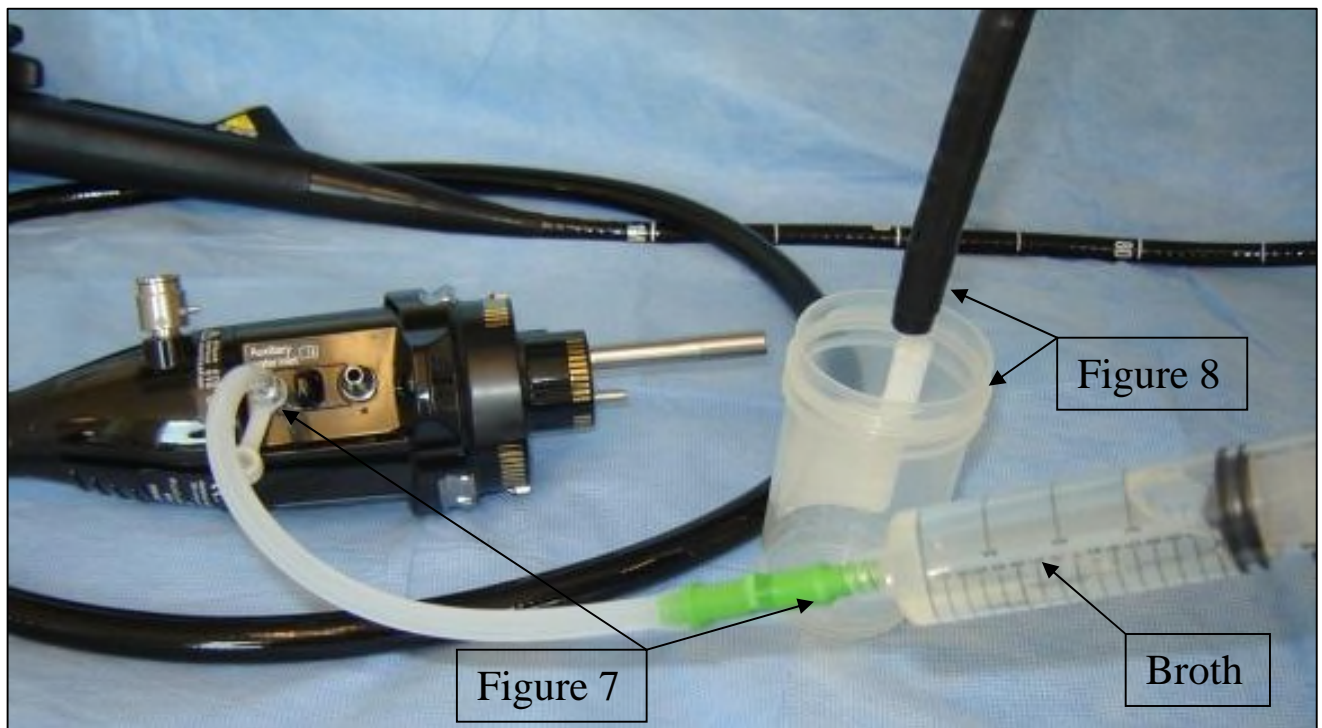


## Step 7 Scope Culture Testing

### Third – Culture the “Auxiliary”

*\*Skip this Step 7 if you are testing a scope that does not have an Auxiliary scope port.*

1. Attach Olympus auxiliary tubing to the auxiliary scope port (Figure 7).
2. Draw up 20ml syringe of broth.
3. Do NOT hold down any buttons.
4. Push the entire 20ml syringe of broth through the Olympus auxiliary tubing, followed by 2 to 3 times - loaded with just air. Hold distal end of the scope, allowing broth to empty into the specimen container (Figure 8).



Auxiliary Tubing

Auxiliary Scope Port



## Step 8 Scope Culture Testing

### Fourth – Flushing the Scope

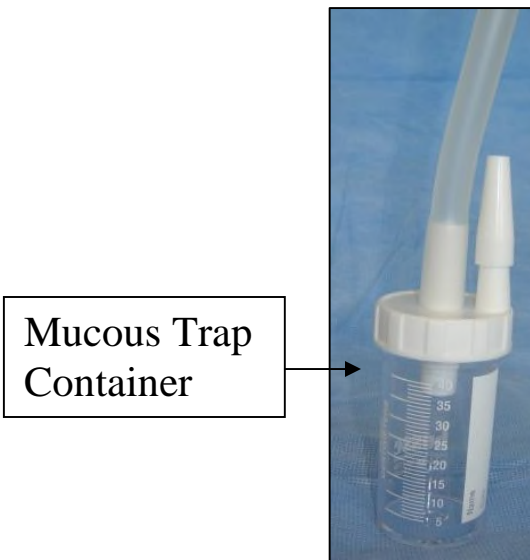
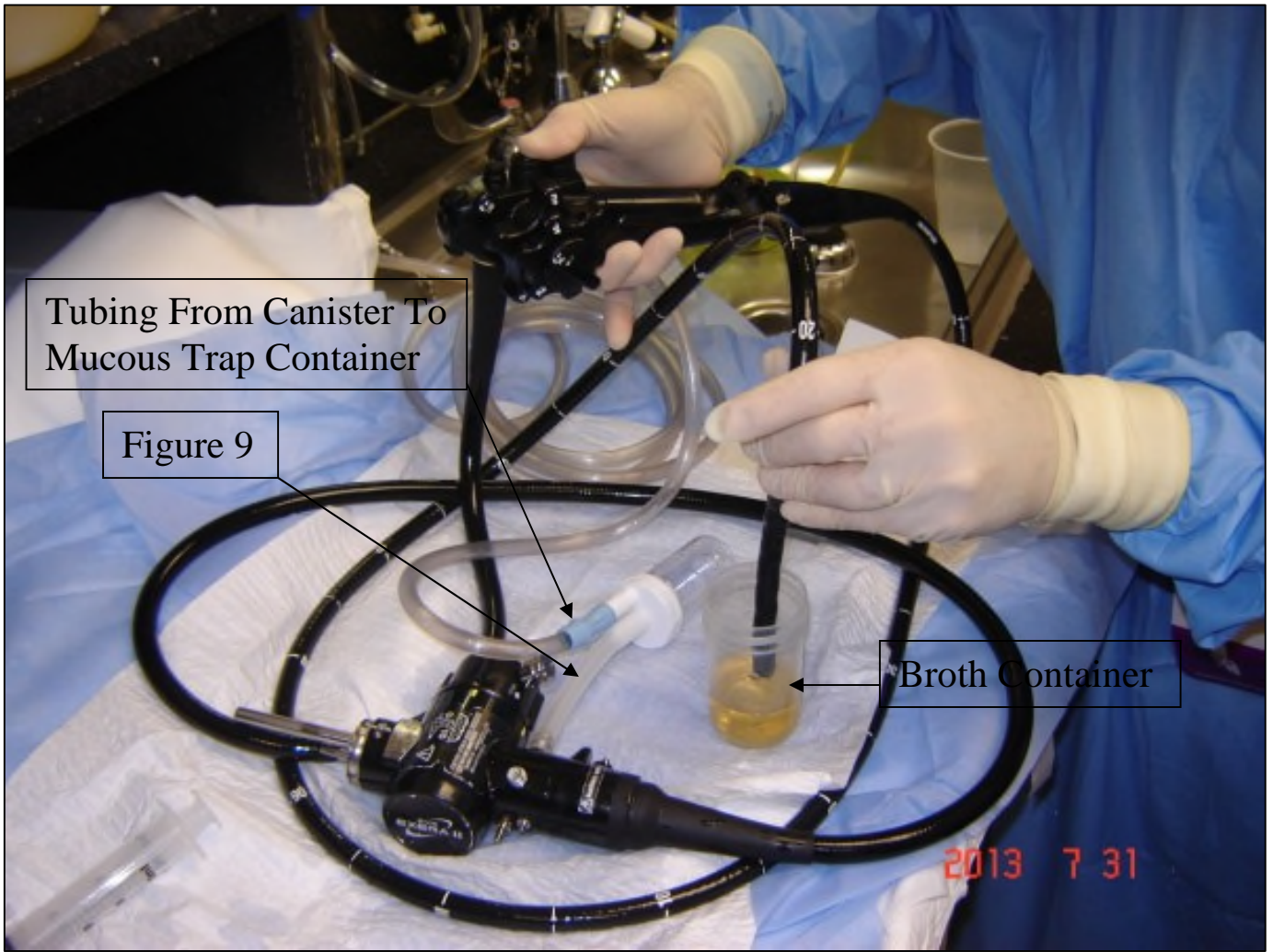
1. Remove all remaining fluid / broth from scope.
  - a. Open suction tubing and place one end on the suction cap on the suction canister.
  - b. Place other end of tubing on the silver suction nozzle of the scope in order to remove any fluid remaining in the scope.

## Step 9 Scope Culture Testing

### Fifth – Culture the “Suction”

1. Then remove the suction tubing from the scope and put the tubing of mucous trap container on silver suction nozzle in order to collect broth once it's ran through the channel.
2. Then connect the previous end of the suction tubing onto the small port of the mucous trap container (Figure 9).
3. Wipe the distal end of the scope with an alcohol wipe and turn on the suction canister. Gently place the distal end into the container without touching the sides, and at the same time you are pushing the **red** suction button only. *Be prepared to quickly disconnect the mucous trap container from the scope or the broth will continue to move into the suction canister.*
4. Turn off the suction canister. Connect rubber tubing of the mucous trap to small port on the trap in order to make it a closed system.





# No Interruptions



## Scope Testing

Place this sign outside your testing facility.

# Cliff Notes

## Step 5 Scope Culture Testing

1<sup>st</sup> – Culture the “Air” Channel

Hold down **red** suction button



## Step 7 Scope Culture Testing

3<sup>rd</sup> – Culture the “Auxiliary”

Do **NOT** hold down any buttons



## Step 6 Scope Culture Testing

2<sup>nd</sup> – Culture the “Water” Channel

Hold down both the **red** suction and the **blue** air and water buttons



## Step 9 Scope Culture Testing

5<sup>th</sup> – Culture the “Suction”

Hold down the **red** suction button



If you have any questions please contact me at [Daniel.keck@unch.unc.edu](mailto:Daniel.keck@unch.unc.edu)