

**Martin S. Favero Lectureship  
Disinfection and Sterilization:  
Successes and Failures**

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Disclosure: Clorox, Advanced Sterilization Products

**Successes**

- Effective cleaning procedures
- Effective and robust high-temperature and low-temperature sterilization technology
- Effective new technologies
  - Low-level disinfection
  - High-level disinfection
  - Prions
- Know how to kill emerging pathogens

**Successes**

**Washer Disinfectors**

**Removal/Inactivation of Inoculum (Exposed) on Instruments**

| WD Conditions | Organism            | Inoculum            | Log Reduction | Positives |
|---------------|---------------------|---------------------|---------------|-----------|
| Routine       | MRSA                | 2.6x10 <sup>7</sup> | Complete      | 0/8       |
| Routine       | VRE                 | 2.6x10 <sup>7</sup> | Complete      | 0/8       |
| Routine       | <i>P aeruginosa</i> | 2.1x10 <sup>7</sup> | Complete      | 0/8       |
| Routine       | <i>M terrae</i>     | 1.4x10 <sup>8</sup> | 7.8           | 2/8       |
| Routine       | GS spores           | 5.3x10 <sup>6</sup> | 4.8           | 11/14     |
| No Enz/Det    | VRE                 | 2.5x10 <sup>7</sup> | Complete      | 0/10      |
| No Enz/Det    | GS spores           | 8.3x10 <sup>6</sup> | 5.5           | 8/10      |

**Sterilization of “Critical Objects”**

Steam sterilization  
Hydrogen peroxide gas plasma  
Ethylene oxide  
Peracetic acid (0.2%)-chemical sterilization  
Ozone  
Vaporized hydrogen peroxide

**High Level Disinfection of  
“Semicritical Objects”**

**Exposure Time ≥ 12 m-30m (US), 20°C**

| Germicide                             | Concentration |
|---------------------------------------|---------------|
| Glutaraldehyde                        | ≥ 2.0%        |
| Ortho-phthalaldehyde (12 m)           | 0.55%         |
| Hydrogen peroxide*                    | 7.5%          |
| Hydrogen peroxide and peracetic acid* | 1.0%/0.08%    |
| Hydrogen peroxide and peracetic acid* | 7.5%/0.23%    |
| Hypochlorite (free chlorine)*         | 650-675 ppm   |
| Accelerated hydrogen peroxide         | 2.0%          |
| Glut and phenol/phenate**             | 1.21%/1.93%   |

\*May cause cosmetic and functional damage; \*\*efficacy not verified

## Low-Level Disinfection for “Noncritical” Objects

| Exposure time $\geq$ 1 min    |                         |
|-------------------------------|-------------------------|
| Germicide                     | Use Concentration       |
| Ethyl or isopropyl alcohol    | 70-90%                  |
| Chlorine                      | 100ppm (1:500 dilution) |
| Phenolic                      | UD                      |
| Iodophor                      | UD                      |
| Quaternary ammonium           | UD                      |
| Accelerated hydrogen peroxide | 0.5%                    |

UD=Manufacturer's recommended use dilution

## Inactivation of Prions

### Recent Studies

- Yan et al. Infect Control Hosp Epidemiol 2004;25:280.
  - Enzymatic cleaner (EC)-no effect
- Fichet et al. Lancet 2004;364:521.
  - Phenolic (Environ LpH), alkaline cleaner (AC), EC+VHP-effective
- Baier et al. J Hosp Infect 2004;57:80. AC-effective
- Lemmer et al. J Gen Virol 2004;85:3805.
  - SDS/NaOH, AC, 0.2% PA, 5% SDS-effective (in vitro)
- Jackson et al. J Gen Virol 2005;86:869. E (Pronase, PK)-effective
- Race R and Raymond G. J Virol 2004;78:2164.
  - Environ LpH-effective
- Peretz et al. J Virol 2006;80:1. Acidic SDS and SDS+SS-effective
- Fichet et al. JHI 2007;67:278. Gaseous HP-effective
- Yan et al. Zentr Steril 2008;16:26-34 HP Gas Plasma effective (Sterrad NX)

## Disinfection and Sterilization of Emerging Pathogens

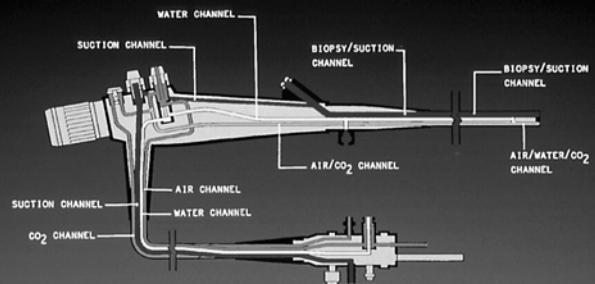
- Hepatitis C virus
- *Clostridium difficile*
- *Cryptosporidium*
- *Helicobacter pylori*
- *E.coli* O157:H7
- Human papilloma virus
- Antibiotic-resistant microbes (MDR-TB, VRE, MRSA)
- SARS Coronavirus, avian/swine influenza, norovirus
- Bioterrorism agents (anthrax, plague, smallpox)

## Failures

## Failures

- Compliance
  - High level disinfection
  - Low level disinfection
    - ◆ Suboptimal surface cleaning/disinfection practices
    - ◆ Disconnect between science and registration process
- Flash Sterilization

## ENDOSCOPE CHANNELS



## Endoscope Reprocessing: Current Status of Cleaning and Disinfection

- Guidelines
  - Centers for Disease Control and Prevention, 2008
  - Multi-Society Guideline, 11 professional organizations, 2003
  - Society of Gastroenterology Nurses and Associates, 2000
  - European Society of Gastrointestinal Endoscopy, 2000
  - British Society of Gastroenterology Endoscopy, 1998
  - Gastroenterological Society of Australia, 1999
  - Gastroenterological Nurses Society of Australia, 1999
  - American Society for Gastrointestinal Endoscopy, 1996
  - Association for Professional in Infection Control and Epidemiology, 2000

## Endoscope Reprocessing, Worldwide

- Worldwide, endoscopy reprocessing varies greatly
  - India, of 133 endoscopy centers, only 1/3 performed even a minimum disinfection (1% glut for 2 min)
  - Brazil, "a high standard ... occur only exceptionally"
  - Western Europe,  $\geq 30\%$  did not adequately disinfect
  - Japan, found "exceedingly poor" disinfection protocols
  - US, 25% of endoscopes revealed >100,000 bacteria

Schembre DB. Gastroint Endoscopy 2000;10:215

## TRANSMISSION OF INFECTION

- Gastrointestinal endoscopy
  - >300 infections transmitted
  - 70% agents *Salmonella sp.* and *P. aeruginosa*
  - Clinical spectrum ranged from colonization to death (~4%)
- Bronchoscopy
  - 90 infections transmitted
  - *M. tuberculosis*, atypical *Mycobacteria*, *P. aeruginosa*

Spach DH et al Ann Intern Med 1993; 118:117-128 and Weber DJ, Rutala WA Gastroint Dis 2002

TABLE 1. Reprocessing Failures of Semicritical or Critical Medical Instruments Resulting in Patient Notification

| Location or institution, year | Instrument involved       | No. of persons exposed |
|-------------------------------|---------------------------|------------------------|
| Sacramento, CA, 2002          | Endoscope                 | 750                    |
| Toronto, ON, 2003             | Endoscope                 | 146                    |
| Seattle, WA, 2004             | Endoscope                 | 600                    |
| Sacramento, CA, 2004          | Endoscope                 | 1,331                  |
| San Francisco, CA, 2004       | Endoscope                 | 2,000                  |
| Long Island, NY, 2004         | Endoscope                 | 177                    |
| Charleston, NC, 2004          | Endoscope                 | 1,383                  |
| Toronto, ON, 2003             | Prostate biopsy probe     | 900                    |
| Pittsburgh, PA, 2005          | Endoscope                 | 200                    |
| Leesburg, VA 2005             | Endoscope                 | 144                    |
| San Diego, CA, 2006           | Endoscope                 | 300                    |
| Augusta, ME, 2006             | Prostate biopsy needle    | 481                    |
| Dept Veterans Affairs, 2006   | Prostate biopsy equipment | 2,075                  |
| San Diego, CA, 2006           | Surgical instrument       | 82                     |

NOTE. Modified from a presentation by Douglas Nelson, MD, at the 33rd Annual Conference and International Meeting of the Association for Professionals in Infection Control and Epidemiology; Tampa, Florida, 2006.

## Disinfection and Sterilization New Systems and Technologies

- New technology that eliminates risk (AERs) or improved compliance
- Elimination of high-level disinfection
  - Improve low-temperature sterilization process so all semicritical items can be sterilized (no restrictions, simple and inexpensive)
  - Develop semicritical items that can be steam sterilized
  - Develop disposable semicritical items (e.g., endoscopes)

## EVOTECH w/Cleaning Claim

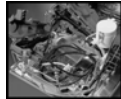


- Product Definition:
  - Integrated double-bay AER
  - Eliminates manual cleaning
  - Uses New High-Level Disinfectant (HLD) with IP protection
  - Single-shot HLD
  - Automated testing of endoscope channels and minimum effective concentration of HLD
  - Incorporates additional features (LAN, LCD display)

## Reliance™ EPS Endoscope Processing System



Reliance™ DG



Endoscope Processing Support



Klenzyme®, CIP® 200



Reliance™ PI

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## Risk of Acquiring MRSA, VRE, and *C. difficile* from Prior Room Occupants

- Admission to a room previously occupied by an MRSA-positive patient or VRE-positive patient significantly increased the odds of acquisition for MRSA and VRE (although this route is a minor contributor to overall transmission). Arch Intern Med 2006;166:1945.
- Prior environmental contamination, whether measured via environmental cultures or prior room occupancy by VRE-colonized patients, increases the risk of acquisition of VRE. Clin Infect Dis 2008;46:678.
- Prior room occupant with CDAD is a significant risk for CDAD acquisition. ICACC (K-4194) 2008. Shaughnessy et al.

## Role of the Environment in Transmission

Pathogens implicated in transmission via contaminated noncritical surfaces (survival in the environment and recovered from the environment)

- Bacteria
  - Oxacillin-resistant *Staphylococcus aureus*
  - Vancomycin-resistant *Enterococcus spp.*
  - *Clostridium difficile*
  - *Acinetobacter* and *P. aeruginosa*
- Viruses
  - Rotavirus
  - Norovirus
  - SARS coronavirus

## Environmental Contamination MRSA

- 27% of 350 surfaces sampled in the rooms of affected patients were contaminated with MRSA. When patients had MRSA in a wound or urine, 36% of surfaces were contaminated. Boyce et al. ICHE 1997;18:622.
- 74% of 359 swabs taken before cleaning yielded MRSA. French et al. J Hosp Infect 2004;57:31

## The Inanimate Environment Can Facilitate Transmission



X represents VRE culture positive sites

~ Contaminated surfaces increase cross-transmission ~  
Abstract: The Risk of Hand and Glove Contamination after Contact with a VRE (+) Patient Environment. Hayden M, ICAAC, 2001, Chicago, IL.

### ***C. difficile* Environmental Contamination**

- Frequency of sites found contaminated ~10->50% from 13 studies-stethoscopes, bed frames/rails, call buttons, sinks, hospital charts, toys, floors, windowsills, commodes, toilets, bedsheets, scales, blood pressure cuffs, phones, door handles, electronic thermometers, flow-control devices for IV catheter, feeding tube equipment, bedpan hoppers
- *C. difficile* spore load is low; 7 studies assessed the spore load and most found <10 colonies on surfaces found to be contaminated. Two studies reported >100; one reported a range of "1->200" and one study sampled several sites with a sponge and found 1,300 colonies *C. difficile*.

## **Practice or Product**

## **Practice\* NOT Product**

\*surfaces not wiped

### **Patient Area Cleaning/Disinfecting**

PC Carling et al, ICHE 2008;29:1 and ICHE 2008;29:1035

- Monitor cleaning performance using an invisible fluorescent targeting method. Rooms (14 high-touch objects) were marked and evaluated after terminal cleaning.
- Results: 1,605 rooms and 20,646 objects were evaluated in 36 hospitals. Mean proportion of objects cleaned was 48%. Following education and process improvement feedback, cleaning improved to 77%
- Conclusion: Substantial opportunity for improving terminal cleaning/disinfecting activities.

**TABLE. Rates of Cleaning for 14 Types of High-Risk Objects**

| Object                | Percentage cleaned |        | 95% CI |
|-----------------------|--------------------|--------|--------|
|                       | Mean ± SD          | Range  |        |
| Sink                  | 82 ± 12            | 57-97  | 77-88  |
| Toilet seat           | 76 ± 18            | 40-98  | 68-84  |
| Tray table            | 77 ± 15            | 53-100 | 71-84  |
| Bedside table         | 64 ± 22            | 23-100 | 54-73  |
| Toilet handle         | 60 ± 22            | 23-89  | 50-69  |
| Side rail             | 60 ± 21            | 25-96  | 51-69  |
| Call box              | 50 ± 19            | 9-90   | 42-58  |
| Telephone             | 49 ± 16            | 18-86  | 42-56  |
| Chair                 | 48 ± 28            | 11-100 | 35-61  |
| Toilet door knobs     | 28 ± 22            | 0-82   | 18-37  |
| Toilet hand hold      | 28 ± 23            | 0-90   | 18-38  |
| Bedpan cleaner        | 25 ± 18            | 0-79   | 17-33  |
| Room door knobs       | 23 ± 19            | 2-73   | 15-31  |
| Bathroom light switch | 20 ± 21            | 0-81   | 11-30  |

NOTE. CI, confidence interval.

**Mean proportion of surfaces disinfected at terminal cleaning is ~50%**

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### Quality Improvement

### Monitoring the Effectiveness of Cleaning

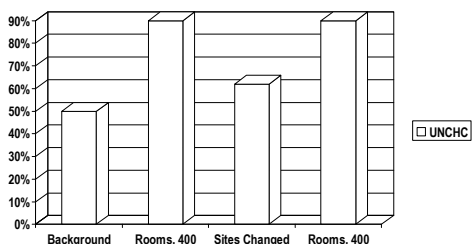
Cooper et al. AJIC 2007;35:338

- Visual assessment-not a reliable indicator of surface cleanliness
- ATP bioluminescence-measures organic debris (each unit has own reading scale)
- Microbiological methods-<2.5CFUs/cm<sup>2</sup>-pass; can be costly and pathogen specific
- Fluorescent marker

### Fluorescent Marker

- A mixture of several glues, soaps, and a target dye (Carling, 2009)
  - Dries rapidly
  - Simple
  - Easily removed by wetted cloth
  - Environmentally stable
  - Rapid
  - Unfortunately, not readily available (Carling and Sodexho)

### Rates of Cleaning for High-Risk Objects



### Room Decontamination Units

MRSA, VRE, *C. difficile*

- Hydrogen peroxide vapor
- Hydrogen peroxide gas
- UV

## Hydrogen Peroxide Vapor Decontamination

- Bartels MD et al. J Hosp Infect 2008;70:35. MRSA/Sterinis
- Boyce JM et al. ICHE 2008;29:723. *C. difficile*/Bioquell
- Shapely S et al. J Hosp Infect 2008 (in press). *C. difficile*/Sterinis
- Hardy KJ et al. J Hosp Infect 2007;66:360. MRSA/Bioquell
- Hall L et al. J Clin Microbiol 2007;45: 810. *M. tuberculosis*/Bioquell
- Bates CJ, Pearse R. J Hosp Infect 2005;61:364. *S. marcescens*/Bioquell
- Johnston MD et al. J Microbiol Methods 2005;60:403. *C. botulinum*/Bioquell
- French GL et al. J Hosp Infect 2004;57:31. MRSA/Bioquell
- Heckert RA et al. Appl Environ Microbiol 1997;63:3916. Viruses/Steris VHP
- Klapes NA et al. Appl Environ Microbiol 1990;56:503. *Bacillus* spores/Prototype HPV generator

## UV Room Decontamination

- Fully automated, self calibrates, activated by hand-held remote
- Room ventilation does not need to be modified
- Uses UV-C (254 nm range) to decontaminate surfaces
- Measures UV reflected from walls, ceilings, floors or other treated areas and calculates the operation time to deliver the programmed lethal dose for pathogens.
- UV sensors determines and targets highly-shadowed areas to deliver measured dose of UV energy
- After UV dose delivered, will power-down and audibly notify the operator
- Reduces colony counts of pathogens by >99.9% within 20 minutes



## Room Decontamination with UV

(Rutala, Gergen, Weber, 2009, Unpublished Results)

| Organism             | Dose Reading (time)             | Log <sub>10</sub> Reduction (10 sites, 5 replicates) |
|----------------------|---------------------------------|--|
| MRSA                 | ~470 mj/cm <sup>2</sup> (~15m)  | 3.91   |
| VRE                  | ~660 mj/cm <sup>2</sup> (~15m)  | 3.36   |
| <i>Acinetobacter</i> | ~630 mj/cm <sup>2</sup> (~14m)  | 3.77   |
| <i>C. difficile</i>  | ~2120 mj/cm <sup>2</sup> (~50m) | 2.67   |

## Failures

- Compliance
  - High level disinfection
  - Low level disinfection
    - ◆ Suboptimal surface cleaning/disinfection practices
    - ◆ Disconnect between science and registration process
- Flash Sterilization

## Contact Time for Surface Disinfection

- CDC guidelines recommends a 1 minute contact time for noncritical surfaces/items. If user selects exposure conditions that differ from label, the user assumes liability and subject to FIFRA.
- Labels on most products registered by EPA specifies a contact time of 10 minutes (some have times of 1-3 minutes)
- Such a long contact time is impractical because dry time 1-3 minutes
- Multiple investigators demonstrated the effectiveness of these disinfectants against bacteria, yeasts, viruses-remedy disconnect

## Flash Sterilization

### Flash Sterilization AORN, CDC Guidelines

- Flash sterilization used for items that must be used immediately
- Acceptable for processing items that cannot be packaged, sterilized and stored before use
- Because of the potential for serious infections, implanted surgical devices should not be flash sterilized unless unavoidable (e.g., orthopedic screws)
- Do not use flash sterilization for reasons of convenience, as an alternative to purchasing additional instrument sets, or to save time

### Flash Sterilization What is the definition?

- In 1942, Underwood defined flash sterilization as 3 minutes at 250°F for instruments when there is an "extreme emergency".
- In 1969, Perkins redefined flash sterilization to the current definition of an unwrapped item at 270°F for 3 minutes in a gravity sterilizer.

### Flash Sterilization

- Flash sterilization principles as defined by Underwood/Perkins and perpetuated by professional organizations are no longer applicable as the longstanding concerns have changed over the past 40 years. Historically, these issues included:
  - Lack of a timely biological indicator to monitor performance (now 1 hr) ;
  - Possibility for contamination of processed items during transportation to the Operating Rooms (containers ensure aseptic delivery to the OR);
  - Sterilization cycle parameters are minimal (extended exposure times) .
- And while no compromise with patient safety can be tolerated, prohibitions and principles regarding flash sterilization should be reassessed by professional organizations.
- Proposal: comply with current recommendations but recommendations should change to define what cycles/conditions are suboptimal.

### Successes

- Effective cleaning procedures
- Effective and robust high-temperature and low-temperature sterilization technology
- Effective new technologies
  - Low-level disinfection
  - High-level disinfection
  - Prions
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**Thank you**

Martin S. Favero Lectureship, 2009